

## The *next generation* – *Monitor Chlorophyll Fluorometer System*



Open daylight dark adaptation module



Closed daylight dark adaptation module

### *Greater capability-*

- *Daylight dark adaptation* allows measurement of  $q_E$  the fast reacting xanthophyll cycle or energy-dependent exciton quenching,  $q_M$  or chloroplast migration,  $q_T$  or state transitions, and  $q_I$  or photoinhibition, at different times of day, and over long periods of time.
- *From 1 to 32 measuring probe heads* are available for each controller, allowing the longer term measurement of larger plant populations and more affordable configurations.
- *10,000  $\mu\text{mol square topped saturation flash}$  or  $F_M'$  correction option according to Loriaux 2013* for more reliable results.
- *Blue light, Red light, and Chlorophyll Content* probes available

# Innovative Design



Open *daylight dark adaptation* module with far red light



Open *daylight dark adaptation* module from the top



Open *daylight dark adaptation* module view from the side



Closed *daylight dark adaptation* module close up



Water proof system controller on pole or tripod with touch screen control, and wireless WiFi antenna.



A Standard Monitor Chlorophyll Fluorometer head without the daylight dark adaptation module



One of the solar panels available with a 12 volt battery box



One of the waterproof junction boxes with fuses for each line, and an LED status light for each line.

# Innovative Design

## Quenching: *New - measuring capability*

*Programable*

*Dark Adaptation*

*...at any time of day*

*...or several times a day*

*...allows quenching relaxation*

$q_E$  - fast acting xanthophyll cycle

$q_M$  - chloroplast migration

$q_T$  - state transitions

$q_I$  - photoinhibition

Kramer's *new* NPQ(T),  $q_E$ (T), &  $q_I$ (T)

Ruban / Murchie protocol - pNPQ &  $qP_d$

*The new monitor fluorometer includes the widest range of quenching protocols available.*

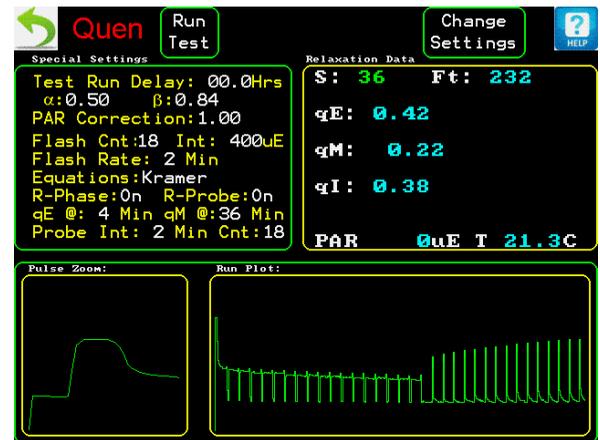
## Hendrickson lake model parameters

Y(II) Quantum yield of PSII

Y(NPQ) Photoprotective non-photochemical quenching

Y(NO) All other non-photo-protective non-photochemical quenching

NPQ Non-photochemical quenching  $NPQ = Y(NPQ) / Y(NO)$



## Kramer lake model quenching parameters (2004)

Y(II) Quantum yield of PSII

$q_L$  Photochemical quenching

Y(NPQ) Photoprotective non-photochemical quenching

Y(NO) All other non-photo-protective non-photochemical quenching

*New Kramer fast quenching parameters are available with the red light version & daylight dark adaptation only NPQ(T),  $q_E$ (T) &  $q_I$ (T)*

## Puddle model quenching

Y(II) Quantum yield of PSII

$q_P$  Photochemical quenching

$q_N$  Non-photochemical quenching

NPQ Non-photochemical quenching

**Chloroplast migration is be responsible for up to about 30% of NPQ at high actinic light levels (Cazzaniga 2013)**

## Quenching relaxation -

$q_E$  Photoprotective energy-dependent exciton quenching - seconds to 7 minutes

$q_M$  Chloroplast migration – chloroplast light avoidance mechanism at high actinic light levels - 20 -35 minutes

$q_Z$  A proposed longer term xanthophyll mechanism – 20 – 30 minutes

$q_T$  The portion of NPQ related to state transitions – 15 to 20 minutes

$q_I$  The portion of NPQ due to photo-inhibition and photodamage. Starts to relax at 40 minutes and may take up to 30 or 60 hours

pNPQ Ruban / Murchie protocol “photoprotective NPQ” designed to measure plant photoprotective mechanisms

$qP_d$  Ruban / Murchie protocol photochemical quenching in the dark immediately after actinic illumination

# Attention to Detail

## ***Algorithm that prevents saturation pulse NPQ issue -***

The instrument measures the highest 25 ms. - 8 point rolling average to determine  $F_M$  and  $F_M'$ . This prevents saturation pulse NPQ from being a problem for all samples, even if the flash width is set too wide. It also eliminates any electronic noise from being a factor.

## ***More reliable leaf temperature measurement -***

Instead of measuring air temperature, the Monitor Fluorometer System offers a leaf thermistor that attaches to the bottom of the leaf for more reliable measurement. *It is completely non-destructive, extremely durable,* and provides more reliable measurement. Measurements are made to  $\pm 0.1^\circ\text{C}$ . Air temperature is also provided from an internal head sensor.

## ***Stable Actinic light source -***

For those instances when one wants to use the internal blue actinic light source, it has been stabilized with a feed-back loop that maintains a constant intensity. Normally, as any light source heats up, the output goes down over time. That does not happen with this instrument. Steady state stable conditions are maintained.

## ***PAR & ETR measurement -***

The instrument measures light intensity at the leaf surface to determine PAR, ETR and when day and night begin.

## ***Saturation flash intensity and $F_M'$ Correction-***

For reliable Y(II) measurements and ETR measurements, the saturation flash of the instrument must be capable closing all PSII reaction centers. If it cannot, an error exists. While this is usually not an issue with dark adapted samples or samples at lower actinic light levels, it is an issue at higher actinic light levels. Research finds that even the most intense saturation flash values cannot completely close all PSII reaction centers. For that reason, the square saturation flash intensity has been increased to  $7,000 \mu\text{mol m}^{-2} \text{ s}$  of blue light, and  $F_M'$  correction, according to Loriaux 2013. Using a red saturation flash, can also be added as an option at time of purchase. Read the section on  $F_M'$  correction for more details. *For those that prefer using a square topped saturation flash, both the blue and red options can provide up to a  $10,000 \mu\text{mol m}^{-2} \text{ s}^{-1}$  saturation flash to minimize this issue.*

## ***Far-red light option -***

Far-red light allows measurement of  $F_O'$ , not just estimations. This allows more reliable Kramer lake model quenching measurements. Far red light is included with the daytime dark adaptation module.  $F_O'$  is estimated when the daytime dark adaptation module is not included.

## ***Easier setup -***

The controller provides a sensitive black and white touch screen interface with drill down menus and graphic displays. Drill down menus provide quick and easy set up, as well as changes. The controller comes with a cover that can be locked. The controller can also be pole mounted for field work.

## ***Add more measuring heads as the need and funding permit -***

One controller can support from 1 to 32 measuring heads, without the added expense of additional controllers. Waterproof junction boxes with fuse lights are available for large numbers of measuring heads and for diagnosis of cabling issues in the field.

## ***Blue or Red light and Chlorophyll Content versions -***

One can choose between blue or red light probe heads, or use some red light units and some blue light units. The same LED is used for modulated light, actinic light and saturating light. Chlorophyll content probe heads, that use the Gitelson ratio fluorescence method, will be available soon.

***Kramer Fast quenching parameters*** - NPQ(T),  $q_E(T)$ , and  $q_I(T)$  are available with the red version head and the daylight dark adaptation option only.

# Attention to Detail

## Interface & Data Management

Data is collected by a single controller and it may be retrieved in various ways. *All systems come with Wi-Fi*, and at time of purchase, investigators can also select other options as needed. They will include cellular modem, Ethernet, Radio point to point, USB stick or satellite phone.

The system interface is a *daylight color touch screen* on the system controller that can be used for programming an data collection. However, *remote system programming and data collection are possible with Smart phones and PCs*. The remote interface is presented as a web page. *Viewing of live measurements* will also be possible.

The measuring files, collected from the controller, are designed to work with Windows Excel (Trademark) or other windows comma delineated programs like Matlab (trade mark). No special computer software is necessary, and as a result, any controller software updates can be done by linking the instrument controller to a PC and downloading updates remotely or by USB stick. The PSP32 offers 2 GB of flash non-volatile memory.

***The monitor fluorometer system has 2 Gb of internal storage that will allow more than a year's worth of data collection, even with 32 measuring heads.***

Rather than providing computer software that must be maintained and updated, the controller will provide information on a computer screens will appear as a web page. Data and graphs can be downloaded and manipulated with other programs, like Excel. The web page data will be password protected.

***For field use, green house, or growth chamber work***

WiFi - comes standard with the controller

### **Options:**

Cellular modem

Ethernet

Radio - point-to-point link available

Satellite modem

USB - stick

# Attention to Detail

Previous research finds that while a high frequency of saturation flashes on light adapted plants will not damage plant tissue, frequent saturation flashes on dark adapted plants, on the same spot, can damage plant tissue. For that reason, the frequency of saturation flashes during the day and night can be programmed separately. Previous research recommends once per hour in the dark.

The measuring trace can be recorded and viewed over very long periods of time. Furthermore, the highest  $F_M$  value during the night, the last  $F_M$  value before dawn, or the highest  $F_M$  value over longer periods of time can be used as the quenching reference. Since it can take from 30 hours to 60 hours for photoinhibition to completely relax or repair, this choice can be valuable depending on study guidelines.

In addition, *The new daylight dark adaptation capability* allows researchers to study  $q_E$  the xanthophyll or cycle energy-dependent exciton quenching,  $q_M$  or chloroplast migration,  $q_T$  or state transitions, and  $q_I$  or photoinhibition, over long periods of time. Here, a fully relaxed  $F_M$  value as described above, can be used with quenching relaxation techniques, during the day, to measure reliable long term results and at different times of day. *The ability to dark adapt during the day allows the use of quenching relaxation protocols required for measuring these very important plant mechanisms.*



We also listened to researchers in regard to *the number of measuring heads* that are available with every controller. To study larger plant populations, the monitor fluorometer system allows up to 32 measuring heads per controller. This keeps the cost lower and allows better science. In addition, daylight dark adaptation modules may be added to *some or all* standard monitor fluorometer measuring heads as money becomes available.

The monitor fluorometer also measures *leaf temperature with a thermistor below the leaf*, not just ambient or instrument temperature. *Instrument or ambient temperature is also measured.*

# $F_M'$ correction – based on Loriaux (2013)

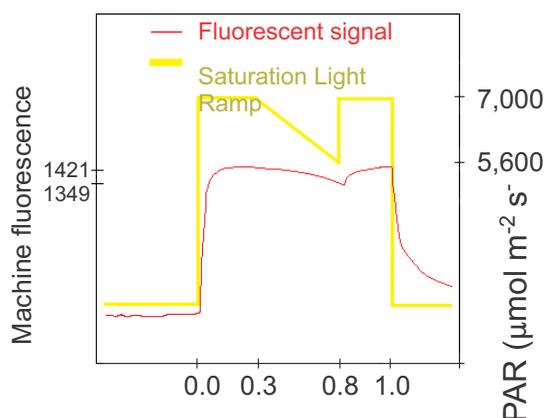
Saturation pulses are used with modulated fluorometers are designed to close all PSII reaction centers. In fact, all reaction centers must be closed to get reliable measurements. The maximum fluorescence intensity value, of the saturation flash called  $F_M'$ , is used in most measurements including, quantum yield of PSII or  $\Phi_{PSII}$  (also called  $Y(II)$  or  $\Delta F / F_M'$ ),  $J$  (or ETR), and in most quenching protocol parameters.

While it is possible to reduce or close all reaction centers in a properly dark adapted sample, with a relatively low amount of light, it has been found that in light adapted samples, *with a high actinic light history*, complete closure of all PSII reaction centers does not occur even with the highest amounts of saturation light. It is thought that complete reduction of  $Q_A$  is prevented by fast turnover of the plastoquinone pools. (Margraph 1990, Loriaux 2013). With this in mind,  $Y(II)$  and ETR measurements, taken under these conditions, can be underestimated. A recent paper (Loriaux 2013), with researchers that included Bernard Genty, the developer of *quantum yield of PSII*, verified the issue, and developed a method for  $F_M'$  correction. It involved a multiple phased single saturation pulse with multiple light intensities, and the use of least squares linear regression analysis of the reciprocal of PAR (Photosynthetically Active Radiation), to determine the  $F_M'$  fluorescence level as if an infinitely intense saturation pulse, were to be used. *This is done without causing damage to the plant and without closing all of the reaction centers.*

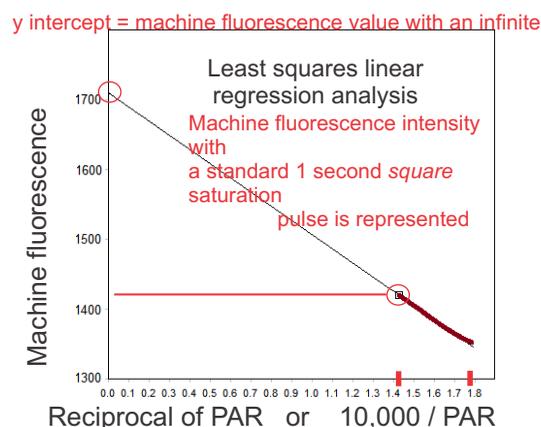
Studies by Loriaux (2006), and Loriaux 2013 have compared chlorophyll fluorescence measurement results with gas exchange measurements and found that a corrected intensity can be determined and used to correct  $\Phi_{PSII}$  or ( $Y(II)$ ) and  $J$  (ETR) measurements. **Research has shown that  $Y(II)$  measurements, taken under high actinic light conditions, can be underestimated with up to a 22% error, and there can be up to a 41% error in ETR values if this method is not used.**

This standard option is provided on the Monitor fluorometer, the  $Y(II)$  meter, the OS5p+, the *iFL*, and the OS1p instruments. It is *available for all Light adapted and quenching protocols*, and it can be turned off or on. The method described in a poster by the Loriaux, Burns, Welles, McDermitt, & Genty (2006) and expanded by Loriaux, Avenson, Welles, McDermitt, Eckles, Riensche, & Genty (2013), provides the most accepted method currently available. According to the science, the Monitor Fluoro meter provides the optimal saturation intensity of 7,000  $\mu\text{mols}$ , optimal light ramping of 20%, and a ramping rate less than 0.01  $\text{mol m}^{-2}\text{s}^{-2}$ . the protocol has been optimized for the best results. The standard monitor fluorometer uses an intense blue saturation light source instead of the red light source in Loriaux 2013. **For those that want to use the standard square topped flash, the instrument offers a 10,000  $\mu\text{mole m}^{-2} \text{s}^{-1}$  blue saturation flash, and the red light version offers a red 10,000  $\mu\text{mole m}^{-2} \text{s}^{-1}$  saturation flash.**

Representation of how the Multiple Phased Flash works



Least squares linear regression of 10,000 / PAR



The first saturation flash step, shown on the left, is at 7,000  $\mu\text{mols}$  for 0.30 seconds to saturate PSII. The saturation flash intensity is then ramped downward by 20%, making a large number of fluorescence measurements along the way, to 5,600  $\mu\text{mols}$ . The ramping rate is less than 0.01  $\text{mol photons m}^{-2}\text{s}^{-2}$ . The final phase is at 7,000  $\mu\text{mols}$  to check for saturation pulse NPQ. Recent studies have shown that those settings provided optimal results for plants that have been tested. (Loriaux 2013). A rolling 25ms eight point average is used to determine maximum  $F_M'$

The graph on the right represents the Loriaux (2013) method for estimating  $F_M'$  with an infinitely intense saturation flash. Least squares linear regression analysis of the reciprocal of PAR (or 10,000 / PAR) allow determination of the y intercept, which represents the machine fluorescence value with an infinite saturation flash.

## Monitor Fluorometer Parameters Measured and Protocols included:

### Light adapted -

**Y(II):** Quantum Yield of PSII

(or )  $\Delta F/F_M'$  or Y)

**ETR:** Electron transport rate

**PAR:** Photosynthetically Active Radiation value

**T:** Leaf temperature

**F<sub>MS</sub> (or F<sub>M'</sub>):** Maximal fluorescence with actinic illumination using a saturation pulse

**F:** Fluorescence under actinic light (prior to saturation pulse)

**Loriaux 2013 correction of ETR, and F<sub>M'</sub> correction** option included for Y(II).

### Dark adapted -

**F<sub>V</sub>/F<sub>M</sub>:** Maximum Photochemical efficiency of PSII

**F<sub>V</sub>/F<sub>O</sub>:** A more sensitive detector of stress than F<sub>V</sub>/F<sub>M</sub>, but it does not measure plant efficiency.

**F<sub>O</sub>:** Minimum fluorescence

**F<sub>M</sub>:** Maximal fluorescence

**F<sub>V</sub>:** Variable fluorescence

**F<sub>O</sub>':** Minimum fluorescence after exposure to far red light- available with the dark adaptation module

**F<sub>O</sub>'** Estimated without the daylight dark adaptation module

With Red light measuring head only - Kramer (2016) fast quenching parameters **NPQ(T)**, **q<sub>E</sub>(T)** & **q<sub>I</sub>(T)**

### Quenching -

**Hendrickson Quenching with NPQ** (standard) **Y(NPQ)**, **Y(NO)**, **Y(II)**, **NPQ**, **F<sub>V</sub>/F<sub>M</sub>**

**Kramer Quenching** (standard)

**q<sub>L</sub>**, **Y(NPQ)**, **Y(NO)**, **Y(II)**, **F<sub>V</sub>/F<sub>M</sub>**

**Puddle model parameters** (standard)

**NPQ**, **q<sub>N</sub>**, **q<sub>P</sub>**, **Y(II)**, **F<sub>V</sub>/F<sub>M</sub>**

**Requires the dark adaptation module option-**

**Quenching relaxation protocol** (option)

**q<sub>E</sub>**, **q<sub>M</sub>**, **q<sub>T</sub>**, **q<sub>Z</sub>**, & **q<sub>I</sub>**. **Ruban / Murchie protocol** **pNPQ** & **qP<sub>d</sub>**

**Rapid Light Curves** (option)

**rETR<sub>MAX</sub>** - ( Eilers and Peeters ) a measure of a leaf's photosynthetic capacity or maximum electron transport rate.

$\alpha$  is the initial slope of line at low PAR values created by relating ETR to PAR. It provides a measure of quantum efficiency

**I<sub>k</sub>** is the measurement of the actinic light intensity where light saturation dominates, or the minimum saturation level.

**I<sub>m</sub>** is the Eilers & Peeters calculated optimal actinic light intensity related to ETR<sub>MAX</sub>

### Light Sources:

**Saturation pulse Blue LED** with:

7,000  $\mu\text{mol m}^{-2} \text{s}^{-1}$  with F<sub>M'</sub> correction option

10,000  $\mu\text{mol m}^{-2} \text{s}^{-1}$  with square topped flash

**Option for Red LED saturation flash**

7,000  $\mu\text{mol m}^{-2} \text{s}^{-1}$  with F<sub>M'</sub> correction option

10,000  $\mu\text{mol m}^{-2} \text{s}^{-1}$  with square topped flash

### Modulated light

Blue 455nm - half band width 18nm

*Option* for red LED 640nm half band width 18 nm.

**Actinic light source:** Blue - Up to 5,000  $\mu\text{mol}$

When the optional red light is used, the actinic light intensity can be up to 5,000  $\mu\text{mol}$

**Far-red light source:** - an option included with the daylight dark adaptation option. It is used to measure F<sub>O'</sub>, or for pre-illumination of samples in the dark adapted mode.

**Detection method:** Pulse modulation method.

**Detector & Filters:** A PIN photodiode with a 700 ~ 750 nm bandpass filter.

**Sampling Rate:** Auto-switching from 1 to 10,000 points per sec., depending on test & on phase of test.

**F<sub>M'</sub> correction according to Loriaux 2013, for all light adapted modes .** It may be turned on or off. Used during daylight hours.

**Test Duration:** Designed to measure samples 24 hours per day, 7 days a week, for months at a time. Runs on solar power, battery power, or mains current.

**Storage Capacity:** 2 Gb. of non-volatile flash memory, supporting almost unlimited data sets and traces. More than 500,000 data sets.

### Special Algorithms:

8 point rolling 25 ms average to determine F<sub>M</sub>, F<sub>M'</sub>, F<sub>O</sub>, & F<sub>S</sub> eliminates saturation pulse NPQ & any electronic noise as an issue.

**Output:** Comma delineated files may be opened in Excel. Data maybe retrieves by WiFi, cell phone, SD data card, Radio point-to-point, Ethernet, satellite phone, or USB stick. Some methods are optional, and require higher pricing.

### User Interface:

*Display: Graphic black and white touch screen interface*  
*Menu driven. Control box may be locked and pole mounted.*

### Power Supply:

Various external 12 volt batteries are available upon request. Solar power and mains power can be used.

**Operating temperature range** -10°C to 50°C

# Accessories

## Optional features & accessories:

- Tripods
- Articulating arms
- Solar panels
- Daylight dark adaptation module with far red light
- Additional measuring heads
- Junction boxes
- Cellular modem
- Radio - point-to-point link available
- Satellite communications
- Stainless 1.5 inch pipe mount with  $\frac{1}{4}$  inch 20 thread mount hole for articulating arms used to mount measuring heads to pole or pipe mounts.
- Integration of other data is possible such as weather station data, humidity, wind speed, infrared radiometer output, soil moisture sensor output, and other radiation sensor output. Contact Opti-Sciences for specific data type and pricing.



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